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10/076,596	02/19/2002	Ronald W. Mink	030793-036100	2706
22204	7590	06/05/2007	EXAMINER	
NIXON PEABODY, LLP 401 9TH STREET, NW SUITE 900 WASHINGTON, DC 20004-2128			NGUYEN, BAO THUY L	
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Please find below and/or attached an Office communication concerning this application or proceeding.

The time period for reply, if any, is set in the attached communication.

Office Action Summary

Application No.

10/076,596

Applicant(s)

MINK ET AL.

Examiner

Bao-Thuy L. Nguyen

Art Unit

1641

-- The MAILING DATE of this communication appears on the cover sheet with the correspondence address --
Period for Reply

A SHORTENED STATUTORY PERIOD FOR REPLY IS SET TO EXPIRE 3 MONTH(S) OR THIRTY (30) DAYS, WHICHEVER IS LONGER, FROM THE MAILING DATE OF THIS COMMUNICATION.

- Extensions of time may be available under the provisions of 37 CFR 1.136(a). In no event, however, may a reply be timely filed after SIX (6) MONTHS from the mailing date of this communication.
- If NO period for reply is specified above, the maximum statutory period will apply and will expire SIX (6) MONTHS from the mailing date of this communication.
- Failure to reply within the set or extended period for reply will, by statute, cause the application to become ABANDONED (35 U.S.C. § 133). Any reply received by the Office later than three months after the mailing date of this communication, even if timely filed, may reduce any earned patent term adjustment. See 37 CFR 1.704(b).

Status

- 1) ☒ Responsive to communication(s) filed on 3/20/2007.
- 2a) ☒ This action is **FINAL**. 2b) ☐ This action is non-final.
- 3) ☐ Since this application is in condition for allowance except for formal matters, prosecution as to the merits is closed in accordance with the practice under *Ex parte Quayle*, 1935 C.D. 11, 453 O.G. 213.

Disposition of Claims

- 4) ☒ Claim(s) 1, 53-55 and 57-71 is/are pending in the application.
- 4a) Of the above claim(s) _____ is/are withdrawn from consideration.
- 5) ☐ Claim(s) _____ is/are allowed.
- 6) ☒ Claim(s) 1, 53-55, 57-71 is/are rejected.
- 7) ☐ Claim(s) _____ is/are objected to.
- 8) ☐ Claim(s) _____ are subject to restriction and/or election requirement.

Application Papers

- 9) ☐ The specification is objected to by the Examiner.
- 10) ☐ The drawing(s) filed on _____ is/are: a) ☐ accepted or b) ☐ objected to by the Examiner.
Applicant may not request that any objection to the drawing(s) be held in abeyance. See 37 CFR 1.85(a).
Replacement drawing sheet(s) including the correction is required if the drawing(s) is objected to. See 37 CFR 1.121(d).
- 11) ☐ The oath or declaration is objected to by the Examiner. Note the attached Office Action or form PTO-152.

Priority under 35 U.S.C. § 119

- 12) ☐ Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f).
- a) ☐ All b) ☐ Some * c) ☐ None of:
1. ☐ Certified copies of the priority documents have been received.
 2. ☐ Certified copies of the priority documents have been received in Application No. _____.
 3. ☐ Copies of the certified copies of the priority documents have been received in this National Stage application from the International Bureau (PCT Rule 17.2(a)).

* See the attached detailed Office action for a list of the certified copies not received.

Attachment(s)

- | | |
|--|---|
| 1) <input type="checkbox"/> Notice of References Cited (PTO-892) | 4) <input type="checkbox"/> Interview Summary (PTO-413)
Paper No(s)/Mail Date. _____ |
| 2) <input type="checkbox"/> Notice of Draftsperson's Patent Drawing Review (PTO-948) | 5) <input type="checkbox"/> Notice of Informal Patent Application |
| 3) <input type="checkbox"/> Information Disclosure Statement(s) (PTO/SB/08)
Paper No(s)/Mail Date _____ | 6) <input type="checkbox"/> Other: _____ |

DETAILED ACTION

1. The amendment submitted 20 March 2007 has been received. Claims 70 and 71 have been added. Claims 1, 53-55, 57-71 are pending.

Claim Rejections - 35 USC § 112

1. The following is a quotation of the second paragraph of 35 U.S.C. 112:

The specification shall conclude with one or more claims particularly pointing out and distinctly claiming the subject matter which the applicant regards as his invention.

2. Claims 57-59 are rejected under 35 U.S.C. 112, second paragraph, as being indefinite for failing to particularly point out and distinctly claim the subject matter which applicant regards as the invention.

Claims 57-59 are indefinite because the analytes cannot be part of the device of claim 56 since these analytes are collected at the time of assay. These analytes are not recited as positive limitation of the device, i.e. they are not reagents that are present on the device nor do they appear to be included in the device prior to the start of the assay.

Claim Rejections - 35 USC § 103

3. The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all obviousness rejections set forth in this Office action:

(a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negated by the manner in which the invention was made.

Art Unit: 1641

4. Claims 1 and 53-71 are rejected under 35 U.S.C. 103(a) as being unpatentable over May (US 5,622,871) in view of Schlipfenbacher (US 5,160,486).

May discloses a device comprising a housing (500) and a strip (506, 510), the strip comprising a collection strip (506) in fluid communication with a lateral flow assay strip (510), wherein the lateral flow assay strip (510) is contained substantially within the housing, contains at least one blocking agent or buffer (see col. 16 line 67 to col. 17 line 40), contains at least one reagent used to detect the presence or absence of an antibody (see col. 16, lines 59-65), contains one or more zones that indicate the presence or absence of the antibody (see col. 19, lines 57-65). The collection strip comprises a capillary matrix adapted for rapid wicking of fluid from a source to the assay strip (see col. 18, lines 35-40). Regarding claim 61, the collection strip protrudes from the housing and is a paddle-shape (see Figs. 8 and 9). The lateral flow assay strip is an immunochromatography strip (see col. 15, lines 4-34). The reagent is a binding partner that bears a detectable label (see col. 15, line 35 to col. 17 line 15). Claims 57-59 fail to further structurally limit the claimed device and instead related to analytes which are not part of the device. These claims discuss analytes, but do not relate the discussion of the analytes to the claimed device. Regarding claims 68 and 69, May ('871) discloses a kit comprising the device discussed above and separately a buffer or reagent (see col. 4, lines 38-42).

May differs from the instant invention in failing to specifically teach a separate blocking strip and a conjugate strip between the collection strip and assay strip,

Schlipfenbacher ('486) teaches providing a blocking strip (23) containing a buffer and a conjugate strip (24) between a collection strip and an assay strip.

It would have been obvious to one of ordinary skill in the art to have provide the blocking strip and conjugate strip between the collection strip and assay strip as taught by Schlipfenbacher ('486) because Schlipfenbacher ('486) expressly teaches providing the strips as an alternative to merely having corresponding separate zones of a single strip (see Fig. 1 vs. Fig. 2).

5. Claims 1, 53-55, 57-58, 60-65, 67-71 are is rejected under 35 U.S.C. 103(a) as being unpatentable over Moorman (US 5,820,826) in view of Ching et al (US 5,120,643).

Moorman discloses a device comprising a capillary matrix (26) having an exposed surface for receiving a test fluid (column 10, lines 22-23). A substrate pad (27) having assay reagents (column 10, lines 23-27), blockings strips including tape and a one-way flow regulating means are located between the capillary matrix and the chromatography strip (column 10, lines 28-32). Moorman teaches that assay reagents may be dried into the pores of the blocking strip. (Column 11, lines 12-21). With respect to claims 63, 65 and 67, Moorman teaches that the apparatus is placed on an inert support or housing (column 5, lines 45-47 and column 11, lines 39-40), which meets the limitation that the chromatography strip extends into the cavity of the housing or the at least partially disposed in the housing. Moorman also teaches that additional features such as antibodies, signal inhibitors, buffers and so forth may be incorporated into the

apparatus. (Column 11, lines 48-51) Moorman teaches that the device is capable of detecting analytes such as HIV antibodies, Rubella, etc. Moorman also teaches kits comprising the device, buffers and reagents for detection of analytes. (Column 26, lines 1-9).

Moorman differ from the instant claims because it fails to disclose that the blocking agents are bovine serum albumin, deoxycholate or n-lauroyl sarcosine.

Ching, however, discloses that devices using labeled specific binding materials including colloidal particle and enzyme labeled materials that are dried onto a chromatographic medium in the presence of a meta-soluble protein are capable of being rapidly resolubilized in the presence of an appropriate solvent such as the sample (column 7, lines 3-10). Ching teaches impregnating solid substrate materials with meta-soluble proteins such as bovine serum albumin and detergents, e.g. sodium deoxycholate, etc. (Column 22, lines 25-47).

Therefore, it would have been obvious to one of ordinary skill in the art at the time the invention was made to add the meta-soluble proteins taught by Ching to the blocking means or substrate pad of Moorman because Moorman teaches that additional features may be incorporated into the apparatus including antibodies, signal inhibitors, buffers and so forth (column 11, lines 49-51) and Ching teaches that improved assay results is achieved using the meta-soluble agents. A skilled artisan would have had a reasonable expectation of success in adding the meta-soluble agents of Ching to the device of Moorman because the addition of agents such as buffers and the like are well

known in the art as indicated by Moorman and the choice of appropriate agents is chosen on the basis of the aim of the assay and the type of the analytes.

Even though Moorman does not specifically disclose oral fluids, this limitation is seen to be an intended use of the claimed device and is not afforded patentable weight. Moorman also does not specifically call the collection matrix "paddle-shaped", however, since the specification does not specifically state the shape or composition of the "paddle-shaped" matrix, only that it has a certain surface area, the collection matrix taught by Moorman is seen to anticipate this limitation.

6. Claim 59 is rejected under 35 U.S.C. 103(a) as being unpatentable over Moorman in view of Ching as applied to claims 1 and 55-57 above, and further in view of Ziegelmaier (US 6,632,628).

See the discussion of Moorman above. Moorman differs from the instant claims in failing to teach the detection of hepatitis. However, Moorman does teach that the analyte and the analyte specific receptors are chosen on the basis of the aim of the assay and discloses typical tests including assays for etiological agents for infectious diseases (column 23, lines 29-38).

Ziegelmaier discloses assays for etiological agents for infectious diseases such as HIV, rubella, hepatitis A and B, etc. See column 4, lines 1-6.

Therefore, it would have been obvious to one of ordinary skill in the art at the time the invention was made to use the device of Moorman to detect analytes such as

hepatitis as taught by Ziegelmaier because Moorman teaches that its device may be used to detect a variety of different analytes including etiological agents for infectious diseases and Ziegelmaier teaches that etiological agents such as hepatitis are well known in the art and can be detected using immunoassays. Furthermore, because Moorman teaches that the analyte and the analyte specific receptors are chosen on the basis of the aim of the assay, one skilled in the art would have had a reasonable expectation of success in using the device of Moorman to detect hepatitis antigens or antibodies as taught by Ziegelmaier.

7. Claims 1, 53, 55, 57-58, 60-61, 63-71 are rejected under 35 U.S.C. 103(a) as being unpatentable over Kremer (US 4,635,488) in view of Sangha (US 5,334,502) and de Zoeten et al (US 5,611,995).

Kremer discloses a sampling device comprising a hollow tube having at least one open end, and a collecting nib secured in the open end of the tube and having an inner extremity facing the interior of the tube and an outer tip projecting beyond the last mentioned end of the tube for contact with a fluid to be collected. The nib comprises a solid, nonfibrous, porous, water-wetable body having porosity sufficient for absorption of the fluid to be collected. The nib is a unitary molded plastic body made of polyethylene or polypropylene and is treated with a wetting agent to impart water-wettability. Column 2, line 48 through column 3, line 9. The device of Kremer is also provided with a cap for closing the open end of the tube and with an elongated,

absorbent and rigid analysis element having an agent that undergoes an observable change upon contact with a substance to be detected in a body fluid sample. The analysis element also has a proximal end mounted in the cap, such that when the cap is in position closing the open end of the tube, the analysis element extends through the tube and its distal end is in fluid transferring contact with the inner extremity of the nib to receive and absorb fluid collected by the nib. The distal end of the analysis element may be anchored in the nib, or may comprise a body of porous material arranged for contact with the inner extremity of the nib so that transfer of the sample from the nib to the analysis element by absorption occurs only after collection of the sample by the nib has been completed. Column 3, lines 26-50. In one embodiment, the device having an analysis element, either in particulate form or strip form, also comprises an absorbent but hydrophobic body situated between the nib and the analysis element to prevent premature transfer of samples to the analysis element (i.e. blocking strip). Column 3, lines 59-65, column 10, lines 53-68, and figure 16. Kremer discloses that the nib absorbs and retains a fluid sample by wicking or capillary action and should be contacted with, for example, the tongue until the nib is completely saturated with the body fluid; since a given porous nib has an essentially fixed fluid capacity, saturation assures collection of a sample of predetermined volume. Column 7, lines 7-23. Kremer discloses that porous nibs may be purchase from Porex Technologies. Column 5, lines 25-28. The device also comprises a transparent sidewall for visual observation of the color change. Column 8, lines 61-66.

Kremer differs from the instant claims in failing to teach that the blocking strip comprises blocking agents and detergents or buffers.

Sangha discloses a method and device for saliva specimen collection comprising a capillary tube surrounding an absorbent pad. On top of the absorbent pad is a one-way barrier having indicator component. As saliva migrates or is wicked along the absorbent pad, it approaches and passes through the one-way barrier to interact with the indicator component. Once the saliva has passed upwardly through the barrier, the saliva is unable to migrate back through the barrier. Thus contact between the subject and the saliva that has interacted with the indicator is avoided. Column 9, lines 37-50. Sangha also discloses a test card for detecting analytes in a saliva sample comprising tetramethylbenzidine (TMB) dissolved in dimethyl formamide (DMF) or dimethyl sulfoxide (DMSO) and EDTA impregnated thereon and dried. Column 15, lines 13-27.

De Zoeten discloses an apparatus having a housing and holding device thereon for holding a test strip comprising a sample collector which can readily absorb test liquid, but also easily release the liquid under capillary transfer. The sample collector is made of material such as polypropylene, and to this material can be added, reagents such as buffering compounds to adjust the pH of the test liquid or compounds able to eliminate interfering substances present in the test liquid. Column 4, lines 40-59. De Zoeten discloses conventional blocking agents such as polyvinylalcohol, or human and bovine serum albumin. Column 7, lines 5-14.

It would have been obvious to one of ordinary skill in the art at the time the invention was made to place reagents such as buffers and detergent taught by Sangha and de Zoeten in the device of Kremer because Sangha and de Zoeten teach that such reagents are well known in the art as providing the advantage of improving assay results by maintaining appropriate pH of the sample and dissolving interference material prior to contacting the sample with the test reagents.

A skilled artisan would have had a reasonable expectation of success in placing these reagents on the blocking strip of Kremer because Sangha teaches a blocking strip made of the same material as that of Kremer which can incorporate reagents such as dyes, and de Zoeten teaches adding buffering compounds to a sample collector (also made of the same material) to adjust pH of the test liquid. Therefore, absent unexpected results, these limitations are seen to be obvious in view of the teachings of Kremer as modified by Sangha and de Zoeten.

8. Claim 62 is rejected under 35 U.S.C. 103(a) as being unpatentable over Kremer in view of Sangha and de Zoeten as applied to claims 1, 53, 55-58, 60-61 and 63-69 above, and further in view of Porex Technologies Catalog, 1995.

See the discussion of Kremer, Sangha and de Zoeten above. These references differ from the instant claim in failing to teach a capillary matrix having an average pore size from about 40 to 250 μ m.

Porex discloses porous plastics available in molded shapes, sheets, rods and tubes having an average pore size from 7 to greater than 250 micrometers. Porex engineers can also develop custom designs for specific use that will take into consideration strength, sample flow, durability and shape. See pages 1, 3, 8 and 24-25.

It would have been obvious to one of ordinary skill in the art at the time the invention was made to choose a porous nib with the desired pore size such as taught by Porex for use in the device of Kremer or Moorman as modified by Sangha and de Zoeten because this parameters are dependent on the nature of the assay, i.e. samples to be tested and reagents involved. Therefore, a skilled artisan would have had a reasonable expectation of success in choosing from any of the disclosed nibs or to have nibs specification made to fit their needs. The selection of a specific material is generally dependent on the assay and the characteristics of the sample, therefore, absent unexpected or improved results, selection of nibs with specific pore sizes so as to optimize the performance of a device is seen to be obvious in view of the teachings of Kremer or Moorman and Porex technologies.

Double Patenting

9. The nonstatutory double patenting rejection is based on a judicially created doctrine grounded in public policy (a policy reflected in the statute) so as to prevent the unjustified or improper timewise extension of the "right to exclude" granted by a patent and to prevent possible harassment by multiple assignees. A nonstatutory obviousness-type double patenting rejection is appropriate where the conflicting claims are not identical, but at least one examined application claim is not patentably distinct from the reference claim(s) because the examined application claim is either anticipated by, or

would have been obvious over, the reference claim(s). See, e.g., *In re Berg*, 140 F.3d 1428, 46 USPQ2d 1226 (Fed. Cir. 1998); *In re Goodman*, 11 F.3d 1046, 29 USPQ2d 2010 (Fed. Cir. 1993); *In re Longi*, 759 F.2d 887, 225 USPQ 645 (Fed. Cir. 1985); *In re Van Ornum*, 686 F.2d 937, 214 USPQ 761 (CCPA 1982); *In re Vogel*, 422 F.2d 438, 164 USPQ 619 (CCPA 1970); and *In re Thorington*, 418 F.2d 528, 163 USPQ 644 (CCPA 1969).

A timely filed terminal disclaimer in compliance with 37 CFR 1.321(c) or 1.321(d) may be used to overcome an actual or provisional rejection based on a nonstatutory double patenting ground provided the conflicting application or patent either is shown to be commonly owned with this application, or claims an invention made as a result of activities undertaken within the scope of a joint research agreement.

Effective January 1, 1994, a registered attorney or agent of record may sign a terminal disclaimer. A terminal disclaimer signed by the assignee must fully comply with 37 CFR 3.73(b).

10. Claims 1, 53, 55-61 and 63-71 are provisionally rejected on the ground of nonstatutory obviousness-type double patenting as being unpatentable over claims 67-80 and 51-53 of copending Application No. 09/973,956. Although the conflicting claims are not identical, they are not patentably distinct from each other because they are both claiming a device for collecting and assay of analytes in oral fluids comprising a housing, a collection pad coupled to a chromatographic strip having appropriate reagents. The device also comprises a blocking strip with blocking agents or buffers.

This is a provisional obviousness-type double patenting rejection because the conflicting claims have not in fact been patented.

Response to Arguments

11. Applicant's arguments filed 20 March 2007 have been fully considered but they are not persuasive.

In response to applicant's arguments against the references individually, one cannot show nonobviousness by attacking references individually where the rejections are based on combinations of references. See *In re Keller*, 642 F.2d 413, 208 USPQ 871 (CCPA 1981); *In re Merck & Co.*, 800 F.2d 1091, 231 USPQ 375 (Fed. Cir. 1986).

Applicant argues that the claimed invention is distinct from May because May neither discloses nor suggests a blocking strip that is impregnated with at least one blocking reagent and/or a conjugate strip that contains lateral flow reagents. For this reasons, Applicant argues that May fails to anticipate and render obvious independently claim 1 and 67.

This argument is not persuasive. May was not cited as a reference that anticipates the claims. Instead, May in view of Schlipfenbacher make obvious the claimed invention. May does teach a chromatographic test strip contained within the housing having at least one blocking agent or buffer (see col. 16 line 67 to col. 17 line 40). The test strip also contains at least one reagent used to detect the presence or absence of an antibody (see col. 16, lines 59-65). May differs in that the buffers and/or blocking agent are not disposed on a separate strip located between the sample receiving pad and the test strip. However, Schlipfenbacher teaches such a buffer strip. Therefore, it would have been obvious to one of ordinary skill in the art to have provide the blocking strip and conjugate strip between the collection strip and assay strip as taught by Schlipfenbacher ('486) because this would have been a mere functionally equivalent means for providing reagents to a test strip. Furthermore, Schlipfenbacher

expressly teaches providing the separate strips as an alternative to merely having corresponding separate zones of a single strip (see Fig. 1 vs. Fig. 2).

Applicant argues that contrary to Schlipfenbacher, the claimed apparatus include a capillary matrix that comprises a porous plastic material while the lateral flow chromatography strip is made from nitrocellulose, whereas the material that comprises the test zones taught by Schlipfenbacher is made of 50% non-swelling fiber fleece materials.

This argument is not persuasive. The features upon which applicant relies (i.e., the specific material of the test strip and the various zones) are not recited in the rejected claim(s). Although the claims are interpreted in light of the specification, limitations from the specification are not read into the claims. See *In re Van Geuns*, 988 F.2d 1181, 26 USPQ2d 1057 (Fed. Cir. 1993). Furthermore, Schlipfenbacher is not cited for their disclosure of the test strip material; it is cited for disclosing that separate reagent impregnated strips are functionally equivalent to test strips in which reagents are impregnated in separate zones.

The recitation that the capillary matrix (i.e. sample receiving zone) is composed of a material different from the material of the lateral flow strip is taught by May.

Applicant argues that Moorman does not teach or suggest a blocking strip containing at least one blocking agent disposed between the capillary matrix and the lateral flow chromatographic strip. Applicant argues that the double-sided tape and the one-way flow regulating means of Moorman are not the same with blocking strip of the

instant claims. Applicant also argues that the blocking strip of Moorman does not have the same function as the claimed blocking strip and that the claims must be interpreted in light of the specification.

This argument is not persuasive. Even though the claims are interpreted in light of the specification, limitations from the specification are not read into the claims. The instant claims do not specifically define the blocking strip and requires only that it contains at least one blocking agent and is disposed between a sample collection matrix and a chromatographic strip. Such a strip is clearly taught by Moorman. Moorman specifically teaches that the double sided tape blocks the flow of the liquid, i.e. a blocking strip, and the one way flow regulating means, in addition to its function as a blocking means, may also contain reagents such as buffers (column 11, lines 12-21 and lines 49-51). Since the claimed "blocking strip" can be broadly interpreted as "to prevent", the blocking assembly taught by Moorman is seen to be equivalent with the claimed blocking strip.

The argument that Moorman teaches away from a capillary matrix having exposed a surface for receiving oral fluid is not persuasive. Clearly, Moorman teaches that liquid sample can be applied to zone 26, the capillary matrix that functions as the sample application area, therefore, this matrix must have an exposed surface, otherwise, fluid cannot be applied to it.

The argument that unlike the absorptive means taught by Moorman, the claimed capillary matrix is not positioned above the lateral flow chromatography strip. Instead,

it lies next to the lateral flow chromatography strip and is in direct communication to the strip. This argument is not persuasive. Nothing in claims 1 or 67 indicates that the capillary matrix lies *next to* the lateral flow chromatography strip. Claims 1 and 67 both recites a capillary matrix extending from within the housing and protruding out from the housing for receiving oral fluid. Moorman teaches such a sample-collecting pad.

The argument that the capillary matrix of the instant invention is made of hydrophilic porous plastic matrix and is different from the sample absorbing means taught by Moorman is not persuasive. This feature of the capillary matrix is not recited in the rejected claims. Again, claims are interpreted in light of the specification, but limitations from the specification are not read into the claims,

The argument that Moorman fails to teach an apparatus for collection and chromatography of oral fluids is not persuasive. Moorman discloses the device essentially as claimed. The intended use of the claimed device is not given patentable weight since it does not structurally change the device.

The argument that Ching fails to disclose or suggest the claimed lateral flow chromatography strip and blocking strip that contains blocking agents and that like Moorman, Ching fails to disclose the claimed apparatus.

This argument is not persuasive. Ching is cited for their explicit teaching of impregnating solid substrate materials with meta-soluble proteins such as bovine serum albumin and detergents, e.g. sodium deoxycholate, etc. (Column 22, lines 25-47). Moorman as discussed above teaches all other limitations of the device. Therefore, a

skilled artisan would have had a reasonable expectation of success in adding the meta-soluble proteins taught by Ching to the blocking means or substrate pad of Moorman because Moorman teaches that additional features may be incorporated into the apparatus including antibodies, signal inhibitors, buffers and so forth (column 11, lines 49-51) and Ching teaches that improved assay results is achieved using the meta-soluble agents.

The argument that Ziegelmaier fails to teach a one-step immunoassay for the determination of antigen-specific antibodies is not persuasive. Ziegelmaier is cited for their teachings of the detection of antibody to hepatitis. Moorman in view of Ching discloses all other limitations of the claimed device.

Applicant argues that in Kremer, the analysis element for sample detection is a removable portion of the sample device and lacks reagents for detecting analytes.

This argument is not persuasive. Nothing in the claims excludes a removal analysis element. Furthermore, the removable analysis element constitutes only one embodiment of the device. The device disclose by Kremer encompasses many other embodiments where the analysis element is not removed from the transparent tube. Kremer discloses specifically at column 8, starting at line 51 an analysis element having an agent that undergoes change of appearance (e.g. color) when it comes into contact with a substance to be detected (i.e. analytes) in a body fluid sample. The strip is constructed of a suitable adsorbent material and constitutes a conventional chromatographic strip containing an agent such that when contacted with analytes in a

sample visual observation of color change provides an indication of the presence or absence (in the sample) of the analyte. In some instances, quantitative analysis may be performed in the same way, e.g. by observing the extent to which a color change develops along the analysis element. In other cases, the active material of the analysis element may undergo changes that are detected by observation with instruments rather than by visual observation.

Clearly, Kremer discloses test strip with at least one reagents used to detect or quantify at least one analyte in an oral fluid sample.

The argument that Kremer does not disclose a blocking strip comprising a blocking agents such as detergents or buffers, and neither Sangha or de Zoeten cure this deficiency is not persuasive. Kremer clearly teaches a blocking strip in the form of a hydrophobic body disposed between the nib (capillary matrix) and the analysis element (chromatography strip) to prevent premature transfer of samples to the analysis element. As stated above, the claimed "blocking strip" can be broadly interpreted as "to prevent", the hydrophobic body taught by Kremer is seen to be equivalent with the claimed blocking strip. Reagents such as buffers and detergents are not disclosed by Kremer as being on the hydrophobic body, however, such reagents are well known in the art as shown on Sangha and de Zoeten.

The argument that Sangha does not teach a blocking pad and blocking agents is not persuasive. Sangha as well as Kremer clearly teach a blocking strip containing reagents such as dyes, and de Zoeten discloses buffering compounds that can

incorporate into a test device such as in the sample collector to adjust the pH of the sample. Therefore, absent unexpected results, these limitations are seen to be obvious in view of the teachings of Kremer as modified by Sangha and de Zoeten.

The argument that the Porex reference does not remedy the deficiencies of Kremer, Moorman, Sangha or de Zoeten is not persuasive. All features of the claimed device are taught Kremer, Moorman, Sangha or de Zoeten as discussed above. Porex is cited for their teachings that porous plastics are widely available in molded shapes, sheets, rods and tubes having an average pore size from 7 to greater than 250 micrometers. Porex engineers can also develop custom designs for specific use that will take into consideration strength, sample flow, durability and shape. See pages 1, 3, 8 and 24-25. Therefore, with the ease of obtaining the appropriate papers, a skilled artisan would have been motivated to choose a porous nib with the desired pore size depending on the nature of the assay, i.e. samples to be tested and reagents involved. A skilled artisan would have had a reasonable expectation of success in choosing from any of the disclosed nibs or to have nibs specification made to fit their needs. The selection of a specific material is generally dependent on the assay and the characteristics of the sample, therefore, absent unexpected or improved results, selection of nibs with specific pore sizes so as to optimize the performance of a device is seen to be obvious in view of the teachings of Kremer or Moorman and Porex technologies.

Conclusion

12. THIS ACTION IS MADE FINAL. Applicant is reminded of the extension of time policy as set forth in 37 CFR 1.136(a).

A shortened statutory period for reply to this final action is set to expire **THREE MONTHS** from the mailing date of this action. In the event a first reply is filed within **TWO MONTHS** of the mailing date of this final action and the advisory action is not mailed until after the end of the **THREE-MONTH** shortened statutory period, then the shortened statutory period will expire on the date the advisory action is mailed, and any extension fee pursuant to 37 CFR 1.136(a) will be calculated from the mailing date of the advisory action. In no event, however, will the statutory period for reply expire later than **SIX MONTHS** from the mailing date of this final action.

13. Any inquiry concerning this communication or earlier communications from the examiner should be directed to Bao-Thuy L. Nguyen whose telephone number is (571) 272-0824. The examiner can normally be reached on Tuesday -- Thursday from 9:00 a.m. - 3:00 p.m..

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Long V. Le can be reached on (571) 272-0823. The fax phone number for the organization where this application or proceeding is assigned is 571-273-8300.

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Bao-Thuy L. Nguyen

Primary Examiner

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